# **REVIEW Open Access**

# Precision diagnosis and treatment of vitamin metabolism-related epilepsy



Yajing Gan<sup>1</sup>, Guoyan Li<sup>1,2</sup>, Zihan Wei<sup>1</sup>, Yan Feng<sup>1,2</sup>, Yuqing Shi<sup>1,2</sup> and Yanchun Deng<sup>1,3\*</sup>❶

# **Abstract**

Epilepsy is a chronic disorder of the nervous system caused by abnormal discharges from brain cells. Structural, infectious, metabolic, immunologic, and unknown causes can contribute to the development of seizures. In recent years, there has been increasing attention on epilepsy caused by genetic metabolic disorders. More than two hundred inherited metabolic disorders have been identifed as potential cause of seizures, and they are mainly associated with energy defciency in the brain, accumulation of toxic substances, abnormal neurotransmitter transmission, and defciency of cofactors. Vitamins play a crucial role as components of several enzymes or coenzymes. Impaired metabolism of thiamine, biotin, vitamin B6, vitamin B12 and folic acid can contribute to early-onset seizures and developmental abnormalities in infants. However, timely supplementation therapy can signifcantly improve patient prognosis of afected patients. Therefore, a thorough understanding and investigation of the metabolic basis of epilepsy is essential for the development of precise therapeutic approaches, which could provide signifcant therapeutic benefts for patients.

**Keywords** Epilepsy, Metabolism, Vitamin, Treatment

# **Background**

Epilepsy is a recurrent and transient disorder of the central nervous system characterized by excessive discharge of nerve impulses in the brain [[1](#page-14-0)]. Previous seizurerelated studies have focused on changes in synaptic transmission. However, as research progresses, disturbances in cellular and mitochondrial metabolism have been recognized as signifcant contributors to the development of epilepsy. Disorders in brain metabolism and seizures may interact, exacerbating the condition. The current global prevalence of inborn errors of metabolism is approximately 1 in 2000 people, with 20–30% of individuals

experiencing epilepsy as their primary neurological manifestation [[2\]](#page-14-1). Oral antiseizure medication therapy is currently the routine treatment for patients with epilepsy, although standardized and appropriate antiseizure medications can control seizures in 70–80% of cases. However, nearly 30% of patients eventually develop drug-refractory epilepsy, which signifcantly afects the quality of life and psychological health of patients, posing a major societal burden [\[1\]](#page-14-0). Among them, mutation-associated epilepsy, in particular, is more likely to progress to drug-resistant epilepsy. Over eight hundred epilepsy-related genes have been identifed, with about 42% of them being metabolically related  $[3]$ . Therefore, understanding and studying the metabolic basis of epilepsy will facilitate the development of more innovative diagnostic and treatment strategies with greater therapeutic benefts.

Currently, genetic metabolic diseases cause epilepsy mainly through mechanisms such as energy deficiency, toxic efects, disturbance of the neurotransmitter system, and cofactor defciency. Among them, vitamins, which serve as coenzymes or coenzymatic components



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/)

<sup>\*</sup>Correspondence:

Yanchun Deng

yanchund@fmmu.edu.cn

<sup>&</sup>lt;sup>1</sup> Department of Neurology, Epilepsy Center of Xijing Hospital, Fourth Military Medical University, Xi'an 710032, People's Republic of China <sup>2</sup> Xi'an Medical University, Xi'an 710021, People's Republic of China

<sup>&</sup>lt;sup>3</sup> Xijing Institute of Epilepsy and Encephalopathy, Xi'an 710000, People's Republic of China

of a variety of enzymes, are not involved in the composition of body tissues and cells or in the production of energy. However, they are mainly responsible for regulating the body's metabolism, producing energy, building and transforming biologically active molecules, and playing an important role in maintaining the function of the nervous system. Vitamin defciencies can therefore lead to a number of systemic metabolic disorders, collectively referred as vitamin defciencies. Defciency in any of the B vitamins can result in neuropsychiatric symptoms. Specifcally, disorders in the metabolism of vitamin B6, thiamine, biotin, vitamin B12, and folic acid metabolism disorders can contribute to drug-resistant seizures and developmental abnormalities in early infancy. Nevertheless, timely treatment with appropriate vitamin supplementation early in life can efectively control seizures and even restore patients to normalcy. Therefore, this review provides a summary of the clinical features and precise treatment protocols for epilepsy related to vitamin metabolism. The aim is to enable clinicians to identify such disorders at an early stage and improve the prognosis of patients.

# **Main text**

# **Vitamin B6 metabolism disorders and epilepsy**

Vitamin B6 encompasses six compounds, namely pyridoxine (PN), pyridoxal (PL), and pyridoxamine (PM), along with their respective 5-phosphates. Of these, pyridoxal 5'-phosphate (PLP) is the most metabolically active form and acts as an enzyme cofactor in the biochemical metabolism of amino acids. Vitamin B6-dependent epilepsy is a group of autosomal recessive disorders caused by mutations in the genes for phosphatidylinositol binding protein (PLPBP), acetaldehyde dehydrogenase 4 family member A1 (ALDH4A1), acetaldehyde dehydrogenase 7 family member A1 (ALDH7A1), pyridoxal (amine) 5'-phosphate oxidase (PNPO), and tissue non-specifc alkaline phosphatase (TNSALP).

In the human body, vitamin B6 is primarily absorbed in the small intestine, where it enters the portal vein and subsequently reaches the liver, then it is converted into PL, PN and PM as well as their respective 5'-phosphates [\[4](#page-14-3)]. A portion of pyridoxine 5'-phosphate (PNP) and pyridoxamine 5'-phosphate(PMP) is oxidized by PNPO to PLP, which is bound to albumin in the blood and travels to all parts of the body. In the central nervous system, PLP is converted to PL across the blood-brain barrier in the presence of TNSALP. Inside brain cells, PL is phosphorylated back to PLP. PLP acts as a universal catalyst and cofactor for amino acids or amines. In the brain, many PLP-dependent enzymes are involved in the metabolism of crucial amino acid or amine neurotransmitters such as dopamine, GABA, glycine, glutamate, and 5-hydroxytryptamine. Therefore, PLP dysfunction can lead to serious central nervous system disorders such as seizures [[5\]](#page-14-4).

# *PLPBP gene mutation and epilepsy*

The *PLPBP* gene, located on chromosome 8p11.23, encodes pyridoxal phosphate homeostasis protein (PLPHP), which is widely expressed in humans, and can be found in the mitochondria or cytoplasm. Its role is to bind PLP and ensure its availability for coenzymes, thus avoiding PLP reacting with other substrates or being phosphorylated by intracellular phosphatases  $[4]$ . Therefore, mutations in the *PLPBP* gene disrupt this process, causing PLP to react with other substrates and impacting the metabolism of important amine or amino acid neurotransmitters in the brain, consequently leading to the development of epilepsy. To date, a total of 54 patients with mutations in the *PLPBP* gene have been reported, with approximately 63% of them have consanguineous parents [[6\]](#page-14-5). Among these cases, 14 patients carried the c.370\_373delGACA mutation, which belongs to the Saguenay-Lac-St-Jean French Canadian founder mutation, in homozygosity and/or compound heterozygosity state. Patients with homozygosity at this mutated locus tend to exhibit a more severe clinical phenotype, usually associated with severe mental retardation and early onset seizures, and five of them died early in life  $[6-8]$  $[6-8]$ . Pregnancy complications, such as preterm birth, fetal distress, and intrauterine hypermobility, were observed in approximately 34% of patients. Almost all preterm infants develop seizures within the frst 24 hours of life, while other affected infants usually develop seizures within the frst week of life. Generalized tonic-clonic (GTC) is the most common seizure type, followed by tonic, clonic, myoclonic, seizures with impaired awareness (SIA), and spasms. In addition to seizures, infantile anemia, infantile gastrointestinal dysfunction, necrotizing small bowel colitis, gastroesophageal refux disease, and metabolic acidosis may occur [\[4\]](#page-14-3). Developmental delay, mental retardation, autism spectrum disorder, acquired microcephaly, and psychiatric abnormalities are also commonly observed in most patients [[4,](#page-14-3) [6,](#page-14-5) [9](#page-14-7), [10](#page-14-8)]. Cranial MRI may reveal simplifed gyri, structural brain abnormalities, periventricular cysts, enlarged ventricles, thinning of the corpus callosum, and high signal in the white matter of the brain.

In individuals with *PLPBP* mutations, the most common metabolic abnormality is an increase in blood lactate and glucose levels, leading to metabolic acidosis shortly after birth, typically on day 1, then returns to normal by day 3 to 4  $[11]$  $[11]$ . Plasma amino acid metabolism is most commonly associated with elevated levels of glycine [\[11\]](#page-14-9). CSF examination also shows elevated



<span id="page-2-0"></span>**Fig. 1** *ALDH4A1* mutation leads to accumulation of P5C in mitochondria. P5C reacts with PLP to form PLP-P5C condensate, which reduces the concentration of PLP in the brain. Elevated P5C can be converted to proline, leading to mitochondrial stress

lactate levels in addition to low homovanillic acid (HVA), along with high levels of 3-O-methyldopa, levodopa, and 5-hydroxytryptophan, suggesting the dysfunction of aromatic L-amino acid decarboxylase (AADC). A study by Akiyama et al. found that reduced CSF PLP concentrations may be a more sensitive indicator of vitamin B6-dependent epilepsy, especially vitamin B6-dependent epilepsy caused by mutations in *PLPBP* [\[12](#page-14-10)]. However, it is important to acknowledge that *PLPBP* still lacks specifc diagnostic features and markers for early detection and prognostic assessment  $[11]$  $[11]$ . Therefore, the diagnosis of PLPHP defciency still relies on standard B6 testing and genetic testing. It is crucial to conduct standard vitamin B6 testing in an environment with full resuscitation capabilities [[13\]](#page-14-11).

Patients with vitamin B6 defciency typically show insensitivity to antiseizure medications but respond well to treatment with PN, and there is a clear dose-dependent relationship [[6](#page-14-5)]. Nearly 91% of patients with *PLPBP* mutations responded to initial treatment with PN and PLP, and about 82% and 50% of patients achieved seizure freedom with PN and PLP, respectively [[8\]](#page-14-6). Long-term maintenance treatment achieves seizure freedom in 69% of patients, with PN in combination with antiseizure medications having the best efficacy. Therefore, it is crucial to start PN treatment as soon as patients are diagnosed with *PLPBP* gene mutations [[14\]](#page-14-12). Switching to PLP may be considered in patients who do not respond well to PN treatment. According to guideline, it is advised that the daily dose of PN should not exceed 200–300 mg in order to prevent PN overdose, which can lead to nerve damage and motor dysfunction [[13,](#page-14-11) [15](#page-14-13)]. Vitamindependent epilepsy requires continuous drug maintenance therapy, and symptoms may return when the drug is discontinued [[14\]](#page-14-12). For patients under 3 years of age with unexplained seizures, it is recommended to undergo intravenous PN testing before starting regular antiseizure medication. Lifelong PN supplementation is initiated if the PN test is positive, and if it is negative, oral PN is continued for several days to avoid overlooking delayed responses.

# *ALDH4A1 gene mutation and epilepsy*

The *ALDH4A1* gene, located at 1p36.13, encodes delta(1)pyrroline-5-carboxylate dehydrogenase (P5CDH). Mutations in this gene can cause type II hyperprolinemia (HPII) with a estimated prevalence of approximately 1 in 700,000 [\[16](#page-14-14)]. The mutation in *ALDH4A1* leads to the accumulation of pyrroline-5-carboxylate (P5C) in the mitochondria, which can react with PLP to cause PLP defciency in the brain and can also be converted to proline, resulting in elevated plasma proline levels (Fig. [1](#page-2-0)) [[17\]](#page-14-15). In patients with hyperprolinemia, proline accumulation can cause a decrease in ATPase activity, which further leads to mitochondrial stress and elevated lactate levels, and can also decrease glutamate uptake [\[17](#page-14-15)]. Thus increased oxidative stress, loss of  $Na^+/K^+$ -ATPase activity, and altered biochemical markers of mitochondrial function play a role in the increased excitotoxicity in individuals with hyperprolinemia.

The most common clinical manifestations are recurrent seizures associated with fever, usually occurring in the neonatal period, early infancy or early childhood. Additionally, individuals may experience a range of developmental delays, mental retardation and signifcant behavioral problems, including schizophrenia, anxiety and hallucinations. Systemic symptoms such as upper respiratory tract infections, vomiting, persistent diarrhea and abdominal pain, respiratory failure, hyperlactatemia and hyperkalemia can also be present [[16,](#page-14-14) [17\]](#page-14-15). However, mild hyperprolinemia is not associated with seizures. So far, a total of eight diferent types of *ALDH4A1* variants



<span id="page-3-0"></span>**Fig. 2** Lysine metabolism. *ALDH7A1* is involved in the oxidation of lysine and mutations in this gene lead to pathological accumulation of α-AASA, P6C and piperonylic acid. P6C accumulation can react with PLP to form a Knoevenagel condensate, which inactivates PLP and interferes with the metabolism of important neurotransmitters in the brain

have been reported, and Kaur et al. reported the frst case of a nonsense mutation leading to premature translation termination [[18\]](#page-14-16).

Lactic acidosis must usually be ruled out before HPII is diagnosed, as it can lead to secondary hyperprolinemia [\[19](#page-14-17)]. Diagnosis of HPII can be made based on plasma proline levels, as well as elevated urinary P5C levels, proline, hydroxyproline, and glycine, which serve as important markers. HPII can be diferentiated from HPI on the basis of urinary P5C levels, but the gold standard for diagnosis is still genetic testing [[19\]](#page-14-17).

Although seizures in patients with *ALDH4A1* mutations may respond to treatment with PN or PLP, these interventions do not improve their intellectual development. According to Hassel et al., the accumulation of γ-glutamate semialdehyde (GSA) and P5C in the brain may contribute to the development of mental retardation in patients [\[20](#page-14-18)]. It has been demonstrated that in animals with *ALDH4A1* mutation, vitamin E and vitamin

C supplementation can reverse the inhibition of ATPase activity by hyperprolinemia, and appropriate use of antioxidants can also be helpful to reduce neurotoxicity [[19](#page-14-17)].

# *ALDH7A1 gene mutation and epilepsy*

The *ALDH7A1* gene, located on chromosome 5q23.2, encodes antiquitin, which is responsible for catalyzing the dehydrogenation of α-aminoadipic semialdehyde (α-AASA)/piperideine 6-carboxylate (P6C). Antiquitin is involved in the oxidation of lysine, and defciency of this enzyme leads to pathological accumulation of α-AASA, P6C, and piperidic acid. In contrast, the accumulation of P6C can react with PLP to form a Knoevenagel condensate, which inactivates PLP and interferes with the metabolism of important neurotransmitters in the brain (Fig. [2\)](#page-3-0) [[21](#page-14-19)]. In particular, it has been suggested that *ALDH7A1* is expressed in glial cells and is involved in early neurogenesis and migration, so mutations in this gene may be associated with abnormal neuronal

migration and structural brain disorders [[22](#page-14-20)]. A total of 165 *ALDH7A1* pathogenic variants have been reported in the literature, but no clear genotype-phenotype correlation has been established. The estimated prevalence of the disease in healthy populations is 1 in 64,352, with a prevalence of 1 in 16,556 in Asia. The most common mutated loci reported in the literature are c.1279G>C and p.Glu427Gln, with 98% of patients having a double allele pathogenic variant [[23](#page-14-21), [24\]](#page-14-22).

Patients with *ALDH7A1* mutations may develop abnormal fetal movements or even intrauterine distress during fetal life. The typical form of seizures occurs within the frst month of life, while in atypical patients, seizures may be delayed until infancy to 3 years of age. Seizures can manifest in a variety of forms, but generalized tonicclonic seizures (GTCS) predominate. Fever can act as a trigger for some patients. In addition to seizures, patients may experience other neuropsychiatric symptoms including autism spectrum disorder, anxiety, attention-deficit/ hyperactivity disorder (ADHD), obsessive-compulsive disorder, mood disorders, hypotonia or hyperactivity, hypervigilance, and sleep disorders. Although treatment with PN may provide partial response, 75% of patients still exhibit residual developmental delays and mental retardation [[25\]](#page-14-23). Systemic symptoms such as vomiting, poor feeding, strabismus, macrocephaly, hypotension, respiratory distress, microcephaly, thrombosis, *E. coli* sepsis, cataracts, hepatomegaly, coagulopathy, uropathy, necrotizing small bowel colitis, jaundice, and hypothermia are less common [\[25](#page-14-23)]. Severe metabolic and endocrine disturbances such as hypocalcemia, hypomagnesemia, hypoglycemia, hypothyroidism, and uroparesis, can also occur in a small number of patients [[24,](#page-14-22) [26](#page-14-24)].

MRI may show structural brain abnormalities such as enlarged ventricles, hypoplasia of the corpus callosum, cortical atrophy, myelin hypoplasia and congenital hydrocephalus, which may be associated with impaired neuronal migration. Brainstem and pontine dysplasia are rare phenotypes, with less than fve patients reported [ $25$ ]. The most common discharge pattern on the EEG is burst suppression, followed by high amplitude dysrhythmias and multi-spike discharges, with corresponding changes in the EEG after the patient receives PN treatment [[27\]](#page-14-25). Plasma threonine, glycine, taurine, histidine, and 3-methoxytyrosine levels are increased in 50% of patients with *ALDH7A1* mutation. CSF levels of GABA, glutamate, threonine, glycine, taurine, histidine, and 3-methoxytyrosine were decreased. Mills et al. analyzed 272 samples and found a significantly higher  $α$ -AASA/ creatinine ratio in this group of patients with a signifcant age dependency  $[24]$  $[24]$ . Urine α-AASA levels should be measured in all newborns with refractory seizures. A signifcant correlation was found between 6R-oxopropylpiperidine-2-carboxylic acid (2-OPP) and *ALDPH7A1* gene defciency in patients. Urinary levels of 2-OPP showed a signifcant positive correlation with α-AASA, and the highest levels of 2-OPP in the CSF were observed in patients treated with vitamin B6. Animal studies suggest that excessive levels of 2-OPP in the brain may lead to persistent neurotoxicity and induce seizures [\[28](#page-14-26)].

Patients with *ALDH4A1* gene mutations typically show poor response to antiseizure medications, while vitamin B6 supplementation has been found to alleviate seizures. However, the response to vitamin B6 may be slower in patients whose epilepsy onset occurs later [[29\]](#page-14-27). Although vitamin therapy can be effective, there may still be a legacy of developmental delays and mental retardation [\[30\]](#page-14-28). Karnebeek et al. discovered that restricting dietary intake of lysine in patients was well tolerated and resulted in a substantial reduction in plasma and CSF levels of α-AASA and P6C, resulting in seizure control and some improvement in developmental milestones [[25\]](#page-14-23). Furthermore, arginine supplementation was found to reduce CSF α-AASA levels and improve motor-verbal function in patients [\[31](#page-14-29)]. Because arginine competitively inhibits lysine transport into the CNS, it reduces the accumulation of P6C and  $α$ -AASA [\[32](#page-14-30)]. Gajlagher et al. found that the *ALDH7A1* mutation is also present in folinic acid-responsive epilepsy, and combined treatment with PN and folinic acid may be considered for patients who do not respond to PN or folinic acid alone [[33\]](#page-14-31). Overall, the main treatment strategy for this group of patients involves vitamin B6 and arginine supplementation to reduce levels of P6C and α-AASA in the brain. Early prevention is also important, and active screening of high-risk individuals and initiation of treatment prenatally may be efective in preventing brain malformations in afected children.

# *PNPO gene mutations and epilepsy*

The *PNPO* gene is situated on chromosome 17q21.32 and encodes pyridoxine-5'-prime-phosphate oxidase, a key enzyme involved in the conversion of PN/PM to PLP. Mutations in this gene can lead to reduced levels of PLP, which can afect the metabolism of several essential amino acids or amine neurotransmitters in the brain [\[34](#page-15-0)].

Mills et al. found that mutations in the *PNPO* genes R225H/C, D33V, and R116Q/P signifcantly reduced *PNPO* activity and increased the likelihood of PNresponsive epilepsy [[35\]](#page-15-1). Individuals with R116Q mutations in the *PNPO* gene have a milder phenotype, with some experiencing no seizures or late-onset symptoms and only mild to moderate mental retardation [\[36](#page-15-2)]. Additionally, most patients with *PNPO* mutations are born prematurely and exhibit abnormal fetal movements, intrauterine distress, and amniotic fuid abnormalities in the fetus. Seizures usually manifest within the frst 2 weeks of life, with a small number of cases may develop after the first year. The epileptic phenotype is diverse, with a predominance of generalized myoclonic seizures, followed by abnormal eye movements and absence sei-zures to a lesser extent [[14](#page-14-12)]. It is worth noting that multisystem abnormalities can also occur, with anemia and coagulopathy being the most frequent hematologic manifestations. Abdominal distension, feeding difficulties, constipation, hepatomegaly, and other gastrointestinal manifestations may also be present  $[14]$ . Approximately 56% of patients have developmental delay and mental retardation. The prognosis is generally poor for those who are born prematurely, have early-onset seizures, and experience delayed treatment  $[37]$  $[37]$ . The EEG shows mainly a burst suppression pattern and completely normal patients are rare  $[14]$  $[14]$  $[14]$ . The most commonly observed fnding on MRI in patients with *PNPO* gene mutations is cerebral atrophy, followed by basal ganglia ischemia, cerebral myelin dysplasia, cortical laminar necrosis and cortical edema. Less commonly, MRI may show intracranial venous sinus thrombosis and subdural hemorrhage [[14\]](#page-14-12). In approximately 80% of patients, elevated levels of glycine can be detected in the CSF, while CSF levels of PLP are decreased in 81% of cases and urinary vanilloid acid is elevated in 91% of patients [[37](#page-15-3)]. Elevated CSF and plasma PM levels and dried blood spot assays showing reduced PNPO enzyme activity are hallmarks of *PNPO* gene mutations [[14\]](#page-14-12).

Vitamin B6 therapy should be started as early as possible in this group of patients, as prompt and timely treatment is strongly associated with a favorable prognosis. Patients who do not receive timely treatment usually have severe developmental delays and severe brain damage. The activity of PNPO is essential for the synthesis of PLP, which is the only theoretically effective therapeutic agent, and must be taken every 6 hours. However, 44% of patients only respond to PN, probably due to residual enzyme activity that converts PN to PLP [[37](#page-15-3)]. It is noteworthy that high concentrations of PLP can regulate intracellular PLP levels by inhibiting the activity of PNPO enzyme, whereas mutant enzymes with residual activity cannot be regulated by PLP negative feedback. As a result, some patients experience worsening symptoms after switching from PN to PLP [\[35\]](#page-15-1). Long-term administration of high doses of PLP can potentially impair liver function, leading to hepatomegaly, cirrhosis, and, in rare cases, hepatocellular carcinoma. Therefore, patients' liver function should be closely monitored, and patients who clearly respond to PN should continue with PN therapy, and the addition of the PNPO cofactor riboflavin may improve the activity of residual PNPO [\[14](#page-14-12)].

# *TNSALP gene mutations and epilepsy*

The *TNSALP* gene, located on chromosome 1p36.12, encodes a tissue non-specifc alkaline phosphatase enzyme, which is widely distributed in the liver, bones, kidneys, and developing teeth. Mutations in the *TNSALP* gene can result in either a complete loss or only 40% of normal alkaline phosphatase activity. However, the degree of reduction in alkaline phosphatase activity is not related to the severity of clinical manifestations [[31\]](#page-14-29). According to the literature, the autosomal recessive inheritance pattern is generally associated with a more severe clinical phenotype compared to autosomal dominant cases [\[38](#page-15-4)]. In 1948, Rathbun frst described the disease as hypophosphatasia in an infant, presented with severe bone loss, seizures, and low levels of alkaline phosphatase tissue and blood, who died of tonic epilepsy at 2 months of age  $[39]$ . The incidence of the disease is approximately 1 in 100,000, and the mortality rate for infantile hypophosphatasia is as high as 50%. In the United States, Whites have shown to be more susceptible to the condition compared to Blacks, and the mortality rate of the disease can further increase when it is associated with infectious atypical pneumonia [\[40](#page-15-6)[–42\]](#page-15-7). *TNSALP* provides inorganic phosphate for the production of hydroxyapatite crystals and promotes the excretion of inorganic pyrophosphate (PPI) [[43](#page-15-8)[–45](#page-15-9)]. The growth of these hydroxyapatite crystals eventually breaks matrix vesicles and mineralizes bone-like material [[46\]](#page-15-10). In contrast, PPI disrupts the growth of hydroxyapatite crystals and inhibits bone mineralization. Therefore, mutations in the *TNSALP* gene result in a decrease in hydroxyapatite production, leading to impaired bone mineralization and decreased bone mass [\[41\]](#page-15-11). In addition, PLP, a cofactor for more than 140 enzymes, is converted to PL by TNSALP across the blood-brain barrier and then converted back to PLP. When the *TNSALP* gene is mutated, it results in low levels of PLP in the CNS, which leads to PN-responsive epilepsy [\[47](#page-15-12), [48\]](#page-15-13).

Based on previous reports in the literature, hypophosphatemia can be classifed into six diferent forms: adult hypophosphatemia, hypophosphatemia in children, infant hypophosphatemia, perinatal hypophosphatemia, benign prenatal hypophosphatemia, and dental phosphate defciency. Among these forms, perinatal hypophosphatemia is the most severe, leading to skeletal damage in the fetus that may not apparent during pregnancy, and often resulting in death shortly after birth. Infantile and perinatal hypophosphatemia are also associated with a variety of acute and chronic systemic manifestations, including poor feeding, anorexia, irritability, persistent vomiting, mild anemia and dehydration, growth retardation, scleral opacities, skin pigmentation, cyanosis, pulmonary infections, hypotony,

and recurrent fractures. Skeletal deformities include cheekbone enlargement, head circumference reduction, cranial stenosis, limb shortening, knee valgus, and, in very severe cases, spherical skulls. Dental abnormalities such as premature loss of primary teeth, hypoplastic teeth, inadequate alveolar bone growth, enlarged pulp spaces, and severe dental caries may also occur. In some cases, hyperphosphatemia and hypercalcemia may even occur due to blocked mineral entry into the bones. Due to abnormal intracranial PLP transport in patients with *TNSALP* mutations, there is insufficient neurotransmitter synthesis in the brain, which eventually contributes to PL-dependent seizures, usually preceding the onset of skeletal changes [[41](#page-15-11), [49–](#page-15-14)[52\]](#page-15-15).

In general, an earlier age of onset of bone disease is associated with a poorer prognosis [\[41](#page-15-11)]. In addition, elevated serum PLP is a sensitive marker for hypophosphatasia, and serum and urine phosphatidylethanolamine levels are usually elevated. Skeletal radiographs are also helpful in diagnosing the disease. Calcitonin and chlorothiazide may be used to treat patients with hypercalcemia and high urinary calcium [\[53](#page-15-16)]. Enzyme replacement therapy for hypophosphatasia has shown promising results in human clinical trials, reducing mortality in hypophosphatasia and improving chondromalacia, lung function and exercise milestones in these patients [\[44](#page-15-17)]. Discontinuing PL supplementation after initiating enzyme replacement therapy can prevent epilepsy recurrence [\[54](#page-15-18)]. Additionally, bone marrow and bone marrow cell transplantation enhances the activity of TNSALP in the bones, and the therapy is currently working well in two female infants as reported [[49,](#page-15-14) [55](#page-15-19)].

# **Thiamine metabolism disorders and epilepsy**

The *SLC19A3* gene is situated at 2q36.3 and encodes thiamine transporter protein 2 (THTR2). *SLC19A1* encodes a reduced folate transporter protein, while *SLC19A2* encodes a thiamine transporter protein. *SLC19A3* shares 39% homology with *SLC19A1* and 48% homology with *SLC19A2*. It is a new heterotrimeric carrier family along with *SLC19A1* and *SLC19A2* transporter proteins, which belongs to the second thiamine transporter and is a potential biotin transporter [\[56](#page-15-20)[–58](#page-15-21)]. When extracellular thiamine concentrations are below 2 mmol/L, active transport via THTR2 is possible. However, at higher extracellular concentrations compared to intracellular concentrations, thiamine enters the cell by passive difusion, so that exogenous thiamin supplementation remains efective even when *SLC19A3* is dysfunctional. In mammals, biotin serves as a coenzyme for several key carboxylases and also regulates several gene expression in vivo. Early biotin defciency inhibits *SLC19A3* expression, but as the defciency worsens, it instead stimulates *SLC19A3* expression and increases biotin transport [\[58](#page-15-21)].

Homozygous and mutated *SLC19A3* gene can cause various conditions such as biotin-thiamine responsive basal ganglia disease (BTBGD), lactic acidosis combined with encephalopathy, infantile epileptic spasms, and early childhood encephalopathy induced by disease or trauma. Among them, mutations such as p.Lys44Glu and p.Glu320Gln in the *SLC19A3* gene can manifest as Wer-Nicke-like encephalopathy [[59](#page-15-22)[–61](#page-15-23)].

Biotin-thiamine-responsive basal ganglia disease, frst described in 1998 by Ozand et al., is an autosomal recessive disorder that is most commonly observed in the Saudi Arabian population. Most parents are consanguineous and more than 100 cases have been reported [[56,](#page-15-20) [60\]](#page-15-24). BTBGD usually occurs in children of preschool or school age and may be precipitated by febrile illness or minor trauma. Clinical symptoms include impaired consciousness, seizures, autism spectrum disorder and depression, as well as extrapyramidal and pyramidal fasciculus symptoms. The patient's intelligence is usually unafected, and in severe cases mild mental retardation may remain. Typical seizure types are simple partial or generalized seizures that can be controlled with one antiseizure medication [\[62](#page-15-25), [63](#page-15-26)]. Extrapyramidal symptoms may manifest as dystonia, dysarthria, and ataxia. Damage to the cone bundle can result in supranuclear facial nerve palsy, extraocular muscle palsy, hemiparesis, and so on  $[63-65]$  $[63-65]$ . Yamada et al. reported two cases of *SLC19A3* pathogenic homozygous mutations (C.958G>C, p.E320Q) with epileptic spasms but atypical EEG manifestations with multifocal spikes and no high-amplitude dysrhythmias [[63\]](#page-15-26). MRI of the brain characteristically reveals centric necrosis of the caudate nucleus head bilaterally and partial or complete necrosis of the shell nucleus in individuals with BTBGD. In addition, there may be supratentorial, infratentorial, and cortical vasogenic edema, with cerebellar, brainstem and thalamic involvement in about 1/3 of patients [\[66,](#page-15-28) [67\]](#page-15-29). Gliosis and cerebral atrophy may occur in patients receiving untimely treatment [\[60\]](#page-15-24). In one case, increased T2 signal intensity was observed in the cervical segment of the spine [\[67](#page-15-29)]. In general, when encountering neurodegenerative disease, epilepsy, or dystonia, together with neuroimaging showing basal ganglia changes should be considered as a possibility of BTBGD [\[68](#page-15-30)].

In 2017, China Xuanwu Hospital proposed the following diagnostic criteria for BTBGD [[69\]](#page-15-31): (i) subacute onset in early childhood; (ii) presence of seizures; (iii) manifestation of dystonia; (iv) display of ataxia; (v) presence of similar symptoms in other family members or children's parents are consanguineous; (vi) identifcation of abnormal signals in the head and shell nuclei of the bilateral

caudate nuclei visible on head MRI; (vii) absence of a better explaination by other diseases; (viii) genetic examination confrming the homozygous or heterozygous mutation of the *SLC19A3* gene. The diagnosis of BTBGD should be highly suspected if the patient meets any two of criteria (i) to (v) and fulfills criteria (vi) and (vii). The diagnosis of BTBGD can be confrmed when the patient's genetic testing meets criteria (viii).

Early treatment is particularly important for patients with BTBGD. Administering high doses of biotin (2–3 mg/kg/d) and thiamine (100–300 mg/d) shortly after symptoms onset can lead to complete resolution or remission of symptoms within a few days in the majority of patients. Failure to receive prompt treatment may result in sequelae such as dystonia, quadriplegia, and mental retardation, and even death. In some patients, using biotin alone may trigger a recurrence of the acute crisis, so biotin is usually combined with thiamine [\[67](#page-15-29)]. Once signifcant remission is achieved through acute treatment, a low-dose maintenance regimen is necessary for long-term management. However, *SLC19A3*-associated infantile epileptic spasms are inefective with large doses of biotin, with only a transient response to ACTH.

# **Biotin metabolism disorders and epilepsy**

Biotinase defciency is caused by mutations in the gene encoding biotinase (*BTD* gene), with 51% of cases attributed to the homozygous c.98-104del7ins3 mutation, It is an autosomal invisible genetic disorder with an estimated prevalence of about 1 in 60,000, and 20% of patients have a history of parental consanguinity [\[70](#page-15-32)]. Mutations in the *BTD* gene decrease biotin activity and reduce the ability of the intestine to absorb biotin, resulting in biotin defciency. In mammals, biotin is a coenzyme of four major carboxylases (pyruvate carboxylase, propionyl coenzyme A carboxylase, 3-methylcrotonyl coenzyme A carboxylase, and acetyl coenzyme A carboxylase). Dysfunction of these four carboxylases can afect gluconeogenesis, lipid synthesis, and amino acid catabolism, leading to multisystem involvement in the body [\[70](#page-15-32), [71\]](#page-15-33).

Most symptomatic children with biotinase defciency exhibit serum biotinase activity at 10% of normal levels or undetectable, and biotin levels in the brain and CSF are typically even lower  $[72]$  $[72]$ . The main manifestations of biotin deficiency are epilepsy, psychomotor retardation, deafness, abnormal muscle tone, skin manifestations, and decreased vision, which usually occur in the frst few months of life. Epilepsy occurs in 55% of patients with the homozygous c.98-104del7ins3 mutation in the *BTD* gene, and for 38% of these patients, epilepsy is the initial symptom  $[73]$  $[73]$ . The most common types of seizures observed are generalized tonic-clonic, followed by myoclonus, epileptic spasms and Ohtahara syndrome [\[72,](#page-15-34) [74](#page-15-36), [75](#page-15-37)]. Skin manifestations include rashes, skin infections, and hair loss, with rashes usually occurring on the face, eyes, mouth, and nose, and sometimes peeling skin [\[75\]](#page-15-37). These skin symptoms may disappear after 1–2 weeks of biotin treatment [\[76](#page-15-38), [77\]](#page-15-39). Metabolic abnormalities afecting the respiratory center of the medulla oblongata may result in hyperventilation, apnea and laryngeal tinnitus. Some patients with biotinase defciency may gradually develop ataxia with age.

The EEG findings in individuals with biotinase deficiency commonly reveal burst suppression and multifocal spikes [\[72](#page-15-34)]. Imaging demonstrates various manifestations, including difuse abnormalities in the supratentorial white matter, reduced brain volume, difuse cerebral edema, basal ganglia calcifcations, and delayed myelination [[78,](#page-15-40) [79\]](#page-15-41). Rarely, bilateral subcortical cysts in the frontal lobe, cranial thickening, and Dandy Walker variant may be observed [[73](#page-15-35)]. MRI scans of three Chinese patients showed the lesions that could involve the spinal cord, with edema and progressive demyelination of the cervical and thoracic spinal cord [\[80](#page-15-42)].

Seizures are controlled by antiseizure medications in about half of patients and disappear after biotin administration, while in patients who do not receive antiseizure medications, most seizures stop within 24 hours of starting biotin application. Some patients with multiple recurrent metabolic disorders may develop irreversible neurological damage with residual mental retardation, ataxia, spastic paraplegia, sensorineural hearing loss, optic nerve atrophy, and epilepsy [\[79](#page-15-41), [81](#page-15-43)]. Because valproic acid impairs mitochondrial function in the liver and reduces biotinidase activity, it should be avoided in this group of patients  $[82]$  $[82]$  $[82]$ . MRI, metabolic screening, and genetic testing should be performed as soon as possible in patients with infantile seizures to establish an early diagnosis and prevent neurologic sequelae with timely and efective treatment [[79](#page-15-41)].

# **Vitamin B12 metabolism disorders and epilepsy**

Vitamin B12 defciency is a common disorder that usually results in megaloblastic anemia, tongue infammation, and neuropsychiatric symptoms  $[83]$  $[83]$ . The most common neurological manifestation is subacute combined degeneration of the spinal cord and peripheral sensory neuropathy, while epilepsy is a rare manifestation in cases of vitamin B12 defciency [\[84](#page-16-2)]. Gramer et al. studied a cohort of 176,702 German children and found 33 children sufered from vitamin B12 defciency with a prevalence of approximately 1:5355 [[85\]](#page-16-3). Irevall et al. screened 35 cases of vitamin B12 deficiency in 11,143 infants in Sweden, with a prevalence of approximately 314 per 100,000 [\[86\]](#page-16-4). Variations in prevalence can be



<span id="page-8-0"></span>**Fig. 3** Metabolic pathways involving vitamin B12. Adenosylcobalamin is involved in the conversion of methylmalonyl coenzyme A to succinyl coenzyme A. In vitamin B12 defciency, methylmalonyl coenzyme A and malonyl coenzyme A accumulate, leading to the synthesis of single-chain fatty acids, which are incorporated into the myelin sheath to cause myelin changes; homocysteine is converted to methionine in the presence of methylcobalamin, methionine is metabolized to S-adenosylmethionine, and S-adenosylmethionine (SAM) is demethylated and converted to S-adenosine homocysteine (SAH). The SAM : SAH ratio decreases in methylcobalamin defciency, protein, lipid and neurotransmitter synthesis in the CNS is impaired, and DNA synthesis and cell division are inhibited

attributed to regional diferences and dietary habits taken into account.

There are currently four theories about the mechanism by which vitamin B12 defciency causes neurological disorders [[87–](#page-16-5)[89](#page-16-6)]: (i) vitamin B12 exists in two active forms, methylcobalamin and adenosylcobalamin. Adenosylcobalamin plays a role in converting of methylmalonyl coenzyme A to succinyl coenzyme A. In vitamin B12 deficiency, the accumulation of methylmalonyl coenzyme A and malonyl coenzyme A, leads to the synthesis of single-chain fatty acids that are incorporated into the nerve sheath, resulting in myelin changes; (ii) homocysteine is converted to methionine in the presence of methylcobalamin, which can be metabolized to S-methionine. However, in methylcobalamin deficiency, the conversion of methionine to S-adenosylhomocysteine (SAH) occurs after the formation of S-adenosylmethionine (SAM), leading to a decrease in the SAM : SAH ratio; (iii) homocysteine accumulation also stimulates N-methyl-Daspartate (NMDA) receptors, leading to the development of neurodegenerative diseases; (iv) elevated levels of TNF- $\alpha$  have also been reported in patients with vitamin B12 defciency, potentially impairing brain development (Fig. [3\)](#page-8-0).

According to current case reports, the phenotype of epilepsy associated with noninfantile vitamin B12 defciency remains uncertain and may present with complex partial seizures, generalized seizures, tonic clonus and generalized tonic-clonic convulsions. Additionally, patients may experience secondary systemic symptoms such as confusion, tendon reflex hyperactivity, reversible dementia, mood swings, psychotic symptoms, and hypertension,. Patients with epilepsy and other neuropsychiatric symptoms who receive cobalamin replacement therapy may resolve or even completely recover [[84,](#page-16-2) [90](#page-16-7)[–93](#page-16-8)]. However, in one case, psychotic symptoms and seizures recurred despite continuous cyanocobalamin replacement therapy after the discontinuation of antipsychotic and antiseizure medications [[90](#page-16-7)].

Patients with infantile vitamin B12 deficiency commonly have vegetarian and pernicious anemic mothers who are also deficient in maternal vitamin B12. These patients usually show developmental delay before the age of 6 months, followed by the onset of seizures. It is

worth noting that all reported cases of vitamin B12-defcient epilepsy in infancy and early childhood present as West syndrome [\[94](#page-16-9)[–96](#page-16-10)]. Meena et al. found that children with infantile spasms had lower mean serum vitamin B12 levels compared to children with generalized developmental delay but without spasms. What's more, children with infantile spasms had higher levels of serum homocysteine and urinary methylmalonic acid compared to the control group, suggesting a potential association between vitamin B12 defciency and the occurrence of infantile spasms [[97\]](#page-16-11). In addition to epilepsy, generalized developmental delay, coma, tachycardia, generalized hypotonia, profound hypotonia, microcephaly, active tendon refexes, and loss of tendon reflexes may be present. A case reported by Pavone et al. described an infant who developed infantile spasms at 8 months of age, as evidenced by high amplitude dysrhythmia on the EEG. Treatment with a combination of vitamin B12, ACTH and vincristine improved neurological function within a few weeks, leading to the disappearance of spasms. However, the patient later developed disseminated bilateral myoclonic seizures with difuse and widespread multi-spike activity on the EEG [\[96](#page-16-10)]. Glaser et al. reported a case involving a 6-month-old infant who was exclusively breastfed and with slowed background activity on the EEG. After starting vitamin B12 supplementation, the patient's cognitive development improved, but clustered spasms developed and the EEG showed hypsarrhythmia [[93\]](#page-16-8). MRI results showed mainly reversible brain atrophy with delayed myelin formation, although some patients had normal MRI fndings [[94–](#page-16-9)[96\]](#page-16-10). In some patients, MRI results may appear normal. Treatment combining with ACTH, antiseizure medications, and vitamin B12 in these patients has shown improvments in EEG features and imaging manifestations, leading to a reduction in seizures. However, there has been a reported case of clinical deterioration with the onset of infantile spasms despite near-normal levels of vitamin B12 in a patient receiving supplementation. Subsequent treatment with topiramate resulted in no further seizures. It suggests that the development of West syndrome in this patient may not be directly related to vitamin B12 defciency [[96\]](#page-16-10). In addition, metabolic analysis may show signifcantly lower serum levels of vitamin B12 and signifcantly higher levels of methylmalonic acid and homocysteine. Whereas the severity of vitamin B12 defciency does not necessarily correlate with prognosis. Attention should be focused on indicators such as methionine and S-adenosylmethionine in plasma and methyl tetrahydrofolate in CSF [[93,](#page-16-8) [95](#page-16-12)].

Overall, vitamin B12 defciency is treatable and can lead to serious neurological sequelae if left untreated, so it is important to consider vitamin B12 defciency as a potential cause in patients with unexplained epilepsy, especially when accompanied by macrocytic anemia [\[85](#page-16-3)]. Long-term use of antiseizure medications is generally discouraged, as medications such as carbamazepine and phenytoin can reduce vitamin B12 and folic acid levels. Hence, it is advisable to avoided these medications [\[98\]](#page-16-13).

# **Folic acid metabolism disorders and epilepsy**

Folic acid is a coenzyme of the one-carbon unit transferase family of biochemical reactions in the human body and is involved in the conversion of homocysteine to methionine and the formation of the active methyl donor, SAM. SAM, in turn, provides methyl for the synthesis of DNA, fatty acids, phospholipids and proteins, with methylation of myelin proteins maintaining the stability of the myelin sheath. The conversion of homocysteine to methionine requires the availability of choline or folic acid to provide methyl groups, so when folic acid is defcient, cells metabolize more choline, resulting in a defciency of phosphatidylcholine, a key component of mycelin. This can lead to a reduction in the stability of the myelin sheath [\[99,](#page-16-14) [100](#page-16-15)].

In 2004, Ramaekers et al. defned primary cerebral folate defciency as a neurological syndrome associated with normal levels of extraneurological folate but reduced levels of 5-methyltetrahydrofolate (5MTHF), a folate metabolite in cerebrospinal fuid [\[101](#page-16-16)]. It may be due to impaired transport of folate into the brain or increased folate metabolism within the brain. Thus, the etiology of primary cerebral folate defciency includes the presence of autoantibodies against the folate receptor (FR1) and mutations in the *FOLR1* gene, which encodes the FR1 protein  $[101]$  $[101]$  $[101]$ . Secondary cerebral folate deficiency encompasses conditions such as dihydrofolate reductase defciency, genetic folate malabsorption, and dihydropteridine reductase deficiency.

# *Primary cerebral folate defciency*

Cerebral folate deficiency is a rare autosomal recessive disorder caused by mutations in the *FOLR1* gene,. To date, 19 cases of cerebral folate defciency due to *FOLR1* gene mutations have been reported. The *FOLR1* gene encodes an adult folate binding protein, which is primarily expressed in the choroid plexus. This protein plays a crucial role in transporting tetrahydrofolate (FH4) across the blood-brain barrier. Patients with cerebral folate defciency usually present with neurological manifestations, mainly ataxia, dystonia, psychomotor hypotonia and seizures after the age of 1 year. Other rare symptoms may include irritability, microcephaly, autism spectrum disorder, tremor, global developmental delay, coma, polyneuropathy, axial hypotonia, and daytime sleepiness [[100,](#page-16-15) [102–](#page-16-17)[108\]](#page-16-18). Among the seizures,

myoclonic and GTCS are the most common, although some patients may experience drop seizures and epileptic spasms. It is important to note that patients in this group usually exhibit normal cognitive function. Brain MRI reveals several abnormalities in patients with cerebral folate defciency, including delayed myelin development, cerebellar atrophy, cortical laminar necrosis, bilateral intrabasal ganglia calcifcations, frontal lobe and cerebellar atrophy. Nuclear magnetic resonance spectroscopy analysis shows reduced peaks of choline and inositol. The impaired function of folate receptor protein 1 (FR1) function due to mutations in the *FOLR1* gene is responsible for these abnormalities. Furthermore, the levels of 5MTHF in the CSF are extremely low or even unmeasurable in this group of patients.

Another important cause of folate deficiency is the presence of autoantibodies against folate receptors in the patient's body, which block the transport of folate across the blood-brain barrier. It typically presents with restlessness, irritability, sleep disturbances initially, which gradually progress to psychomotor retardation, seizures, and cerebellar ataxia [[101](#page-16-16), [109](#page-16-19)]. The disease begins with restlessness, anger, and sleep disturbances. Some cases also present with coma, visual impairment, progressive acoustic hearing loss and spastic paraplegia [[101,](#page-16-16) [109\]](#page-16-19). Similar to the *FOLR1* gene mutations, the most common seizure types observed in this condition are myoclonic and GTCS [[101](#page-16-16)]. Neuroimaging studies may show frontotemporal atrophy, progressive supratentorial and infratentorial atrophy, cerebellar atrophy, and periventricular demyelination [[101](#page-16-16)]. Magnetic resonance spectroscopy analysis often shows decreased levels of N-acetylaspartate [\[109\]](#page-16-19). In addition, the concentration of 5MTHF in the CSF is lower than normal in patients with idiopathic cerebral folate deficiency, and some patients also have reduced concentrations of CSF 5-hydroxyindoleacetic acid and biopterin.

Early initiation of folic acid supplementation (2–10 mg/kg/day) is recommended, and patients who begin supplementation before the age of 6 years tend to have a better prognosis than those who start supplementation after the age of 6 years [\[101](#page-16-16)]. Even if folic acid supplementation is commenced in adulthood, it can still help reduce the frequency of seizures and improve the patient's quality of life. The combination of oral folic acid with intravenous administration is considered more efective than oral administration alone [\[110](#page-16-20)]. However, there are cases where patients experience ongoing deterioration in their developmental status, even after initiating treatment with folic acid and various antiseizure medications at the age of 1 year, with seizures reaching more than 30 times per day  $[110]$  $[110]$ . In addition, the ketogenic diet has a better response when used as an additional treatment along with antiseizure medications [[111\]](#page-16-21). CSF folic acid concentration should be monitored during folic acid supplementation therapy to prevent deficiency or overdose.

# *Secondary folic acid defciency*

The molecular basis of the inherited folate absorption disorder is a mutation in the *SLC46A1* gene, which encodes the proton-coupled folate transporter (PCFT). This transporter is responsible for absorption of folate in the intestine [\[112,](#page-16-22) [113](#page-16-23)]. In addition, patients with hereditary folate absorption disorders also have a folate transporter disorder in the central nervous system, suggesting that PCFT is also involved in the transport of folate to the brain [\[112\]](#page-16-22). Hereditary cerebral folate malabsorption is a secondary cause of cerebral folate malabsorption, with nearly half of the patients having a consanguineous parent and a mortality rate of 40%, with women being more susceptible than men  $[114]$  $[114]$ . The condition primarily manifests as megaloblastic anemia with recurrent diarrhea, mouth ulcers, recurrent infections, anemia and loss of appetite, which typically occur in the frst months of life. Neurological manifestations include developmental delay, psychomotor retardation, behavioral abnormalities, ataxia, peripheral neuropathy, and seizures. Psychiatric symptoms such as insomnia, forgetfulness, irritability, depression, and schizophrenia may also occur [[114\]](#page-16-24). These patients typically have low levels of folate in both plasma and CSF, and they may exhibit abnormally high urinary metformin glutamate excretion [\[115](#page-16-25)]. MRI scan may show calcifcations in the basal ganglia region, occipital cortex, and internal capsule sites. For patients with impaired folic acid absorption, intramuscular folic acid injection may be considered as an alternative to bypass gastrointestinal absorption. If patients continue to have seizures despite folic acid supplementation, additional antiseizure medicatoins should be considered.

The *DHFR* gene is located at 5q14.1 and encodes dihydrofolate reductase. Dihydrofolate reductase is a key enzyme in folate metabolism in vivo, responsible for the reduction of dihydrofolate to tetrahydrofolate, which is involved in the methyl shuttle required for the ab initio synthesis of thymidylate and certain amino acids. Additionally, it serves as a cofactor for enzymes such as phenylalanine 4-hydroxylase, tyrosine 3-hydroxylase, and tryptophan 5-hydroxylase, which are required for the production of monoamines. Only six cases of dihydrofolate reductase defciency have been reported to date, three of which were homozygous mutations in the *DHFR* gene p.Asp153Val and three were attributed to p.Leu80Phe mutations  $[116, 117]$  $[116, 117]$  $[116, 117]$ . The main symptom remains macrocytic anemia, which can be completely



<span id="page-11-0"></span>





reversed by folic acid supplementation. Compared to patients with primary folate defciency, those with dihydrofolate reductase defciency typically exhibit less pronounced developmental abnormalities. However, they may experience respiratory infections, low intraocular pressure, microcephaly, ocular abnormalities, and seizures. The most common type of seizure observed is eyelid myoclonus with absence [\[117](#page-16-27)]. Additionally, strabismus and impaired vision may also be present [\[117](#page-16-27)]. CSF levels of 5-MTHF and tetrahydrobiopterin are lower than normal, while serum levels of folate, dihydrobiopterin, and homocysteine remain within the normal range. There is a slight decrease in the levels of monoamine metabolites such as homovanillic acid (the major catecholamine metabolite) and 5-hydroxyindoleacetic acid (5-hydroxytryptamine metabolite) [[117](#page-16-27)]. MRI may show cerebral white matter atrophy, ventricular enlargement, periventricular gliosis, optic nerve atrophy, delayed myelin formation or dysplasia, cerebellar hypoplasia, cerebellar hemisphere atrophy, and hypersignal in the subcortical white matter [[116](#page-16-26), [117\]](#page-16-27). Folic acid supplementation can alleviate the symptoms of anemia in patients with dihydrofolate reductase defciency, but this does not correct the defciency of tetrahydrofolate in the brain, often leaving neurological sequelae. Furthermore, dihydrobiopterin plays a role in maintaining the reduced state of tetrahydrofolate, so the clinical phenotype of dihydrobiopterin defciency is similar to that of dihydrofolate reductase defciency. However, patients with dihydrobiopterin defciency may develop early brain atrophy and intracranial perivascular calcifcations [[118,](#page-16-28) [119\]](#page-16-29).

3-phosphoglycerate dehydrogenase (3-PGDH) defciency is a rare autosomal recessive disorder of serine synthesis that has been reported in fewer than 20 patients. Deficiency of the 3-PGDH enzyme results in decreased serine synthesis in the body. Serine normally reacts with tetrahydrofolate to form glycine and methylenetetrahydrofolate, which is eventually reduced to 5-methyltetrahydrofolate  $[120]$  $[120]$ . Thus, impaired serine synthesis eventually leads to folic acid defciency. Patients in this group usually present with microcephaly, severe psychomotor retardation, and spastic quadriplegia. Seizures usually occur within the frst 2 years of life, and about half of the afected individuals present with West syndrome [[121\]](#page-16-31). Previous reports have suggested that oral supplementation of serine can improve seizures. However, a study investigated the efects of L-serine supplementation in two patients with PGDH defciency and found that it did not efectively control seizures. Interestingly, with glycine was added to the treatment regimen, seizures were completely controlled, and abnormal brain discharges in the patients showed improvement [\[120,](#page-16-30) [122\]](#page-16-32).

# **Conclusions**

In conclusion, vitamin metabolism disorder-related epilepsy is a type of treatable metabolic epilepsy (Table [1](#page-11-0)). For patients with unexplained mental and motor developmental delay, seizures, recurrent acute encephalopathy, and systemic multisystem manifestations, it is important to conduct early diagnostic investigations such as cranial MRI, EEG, plasma amino acid metabolism, urine organic acid metabolism analysis, CSF metabolism, and genetic testing to clarify the diagnosis. Supplementation with pyridoxine, vitamin B12 and folic acid may be considered. It is advisable to avoid valproic acid, as it can impair liver mitochondrial function and cause hepatotoxic, while carbamazepine and phenytoin can decrease vitamin B12 and folic acid levels, therefore, these drugs should be avoided whenever possible. Levetiracetam and broad-spectrum antiseizure medications such as benzodiazepine may be chosen for treatment. In addition, early prevention is equally important, and individuals at risk should receive appropriate vitamin supplementation during pregnancy. It is expected that further research into the underlying metabolism mechanisms of epilepsy will lead to the development of more innovative treatment strategies, the discovery of more treatable types of epilepsy and greater therapeutic beneft.

#### **Abbreviations**



#### **Acknowledgements**

Not applicable.

# **Authors' contributions**

YG completed the collection and analysis of the relevant literature. YG and YF wrote the draft of the manuscript. ZW, YS, CZ, and GL participated in the analysis and organization of the literature. YD concepted the topic,supervised the project and guided the writing of the manuscript. All authors contributed to the article and approved the submitted version.

### **Funding**

The study was funded by the National Key R&D Program of China (2022YFC2503801).

### **Availability of data and materials**

Not applicable.

#### **Declarations**

**Ethics approval and consent to participate** Not applicable.

# **Consent for publication**

Not applicable.

#### **Competing interests**

Author Yanchun Deng is the member of the Editorial Board for Acta Epileptologica, who was not involved in the journal's review of or decisions related to this manuscript.

Received: 18 February 2024 Accepted: 23 June 2024

#### <span id="page-14-0"></span>**References**

- 1. Devinsky O, Vezzani A, O'Brien TJ, Jette N, Schefer IE, de Curtis M, et al. Epilepsy. Nat Rev Dis Primer. 2018;4:18024.
- <span id="page-14-1"></span>2. Campistol J. Epilepsy in inborn errors of metabolism with therapeutic options. Semin Pediatr Neurol. 2016;23(4):321–31.
- <span id="page-14-2"></span>3. Tumienė B, Peterlin B, Maver A, Utkus A. Contemporary scope of inborn errors of metabolism involving epilepsy or seizures. Metab Brain Dis. 2018;33(6):1781–6.
- <span id="page-14-3"></span>4. Darin N, Reid E, Prunetti L, Samuelsson L, Husain RA, Wilson M, et al. Mutations in PROSC disrupt cellular pyridoxal phosphate homeostasis and cause vitamin-B6-dependent epilepsy. Am J Hum Genet. 2016;99(6):1325–37.
- <span id="page-14-4"></span>5. Wang HS, Kuo MF. Vitamin B6 related epilepsy during childhood. Chang Gung Med J. 2007;30(5):396–401.
- <span id="page-14-5"></span>6. Johnstone DL, Al-Shekaili HH, Tarailo-Graovac M, Wolf NI, Ivy AS, Demarest S, et al. PLPHP defciency: clinical, genetic, biochemical, and mechanistic insights. Brain J Neurol. 2019;142(3):542–59.
- 7. Pal M, Lace B, Labrie Y, Lafamme N, Rioux N, Setty ST, et al. A founder mutation in the PLPBP gene in families from Saguenay-Lac-St-Jean region afected by a pyridoxine-dependent epilepsy. JIMD Rep. 2021;59(1):32–41.
- <span id="page-14-6"></span>8. Alsubhi S, Osterman B, Chrestian N, Dubeau F, Buhas D, Srour M. Case report: PLPHP defciency, a rare but important cause of B6-responsive disorders: a report of three novel individuals and review of 51 cases. Front Neurol. 2022;13:913652.
- <span id="page-14-7"></span>9. Ahmed S, DeBerardinis RJ, Ni M, Afroze B. Vitamin B6-dependent epilepsy due to pyridoxal phosphate-binding protein (PLPBP) defect - First case report from Pakistan and review of literature. Ann Med Surg. 2012;2020(60):721–7.
- <span id="page-14-8"></span>10. Plecko B, Zweier M, Begemann A, Mathis D, Schmitt B, Striano P, et al. Confrmation of mutations in PROSC as a novel cause of vitamin B 6 -dependent epilepsy. J Med Genet. 2017;54(12):809–14.
- <span id="page-14-9"></span>Jensen KV, Frid M, Stödberg T, Barbaro M, Wedell A, Christensen M, et al. Diagnostic pitfalls in vitamin B6-dependent epilepsy caused by mutations in the PLPBP gene. JIMD Rep. 2019;50(1):1–8.
- <span id="page-14-10"></span>12. Akiyama M, Akiyama T, Kanamaru K, Kuribayashi M, Tada H, Shiokawa T, et al. Determination of CSF 5-methyltetrahydrofolate in children and its application for defects of folate transport and metabolism. Clin Chim Acta. 2016;460:120–5.
- <span id="page-14-11"></span>13. Stockler S, Plecko B, Gospe SM, Coulter-Mackie M, Connolly M, van Karnebeek C, et al. Pyridoxine dependent epilepsy and antiquitin defciency: clinical and molecular characteristics and recommendations for diagnosis, treatment and follow-up. Mol Genet Metab. 2011;104(1–2):48–60.
- <span id="page-14-12"></span>14. Wilson MP, Plecko B, Mills PB, Clayton PT. Disorders afecting vitamin B6 metabolism. J Inherit Metab Dis. 2019;42(4):629–46.
- <span id="page-14-13"></span>15. Phillips WE, Mills JH, Charbonneau SM, Tryphonas L, Hatina GV, Zawidzka Z, et al. Subacute toxicity of pyridoxine hydrochloride in the beagle dog. Toxicol Appl Pharmacol. 1978;44(2):323–33.
- <span id="page-14-14"></span>16. van de Ven S, Gardeitchik T, Kouwenberg D, Kluijtmans L, Wevers R, Morava E. Long-term clinical outcome, therapy and mild mitochondrial dysfunction in hyperprolinemia. J Inherit Metab Dis. 2014;37(3):383–90.
- <span id="page-14-15"></span>17. Flynn MP, Martin MC, Moore PT, Stafford JA, Fleming GA, Phang JM. Type II hyperprolinaemia in a pedigree of Irish travellers (nomads). Arch Dis Child. 1989;64(12):1699–707.
- <span id="page-14-16"></span>18. Kaur R, Attri SV, Saini AG, Sankhyan N. A high frequency and geographical distribution of MMACHC R132\* mutation in children with cobalamin C defect. Amino Acids. 2021;53(2):253–64.
- <span id="page-14-17"></span>19. Mitsubuchi H, Nakamura K, Matsumoto S, Endo F. Biochemical and clinical features of hereditary hyperprolinemia. Pediatr Int. 2014;56(4):492–6.
- <span id="page-14-18"></span>20. Hassel B, Rogne AG, Hope S. Intellectual disability associated with pyridoxine-responsive epilepsies: the need to protect cognitive development. Front Psychiatry. 2019;10:116.
- <span id="page-14-19"></span>21. Mills PB, Struys E, Jakobs C, Plecko B, Baxter P, Baumgartner M, et al. Mutations in antiquitin in individuals with pyridoxine-dependent seizures. Nat Med. 2006;12(3):307–9.
- <span id="page-14-20"></span>22. Jansen LA, Hevner RF, Roden WH, Hahn SH, Jung S, Gospe SM. Glial localization of antiquitin: implications for pyridoxine-dependent epilepsy. Ann Neurol. 2014;75(1):22–32.
- <span id="page-14-21"></span>23. Coughlin CR, Tseng LA, Abdenur JE, Ashmore C, Boemer F, Bok LA, et al. Consensus guidelines for the diagnosis and management of pyridoxine-dependent epilepsy due to α-aminoadipic semialdehyde dehydrogenase defciency. J Inherit Metab Dis. 2021;44(1):178–92.
- <span id="page-14-22"></span>24. Mills PB, Footitt EJ, Mills KA, Tuschl K, Aylett S, Varadkar S, et al. Genotypic and phenotypic spectrum of pyridoxine-dependent epilepsy (ALDH7A1 defciency). Brain J Neurol. 2010;133(Pt 7):2148–59.
- <span id="page-14-23"></span>25. van Karnebeek CDM, Tiebout SA, Niermeijer J, Poll-The BT, Ghani A, Coughlin CR, et al. Pyridoxine-dependent epilepsy: an expanding clinical spectrum. Pediatr Neurol. 2016;59:6–12.
- <span id="page-14-24"></span>26. Lauras B, Drevon B, Rolland MO, Teyssier G, Bovier-Lapierre M, Freycon F. Pyridoxine-dependent convulsions : familial case. Pediatrie. 1984;39(3):183–7.
- <span id="page-14-25"></span>27. Nabbout R, Soufflet C, Plouin P, Dulac O. Pyridoxine dependent epilepsy: a suggestive electroclinical pattern. Arch Dis Child Fetal Neonatal Ed. 1999;81(2):F125-129.
- <span id="page-14-26"></span>28. Engelke UF, van Outersterp RE, Merx J, van Geenen FA, van Rooij A, Berden G, et al. Untargeted metabolomics and infrared ion spectroscopy identify biomarkers for pyridoxine-dependent epilepsy. J Clin Invest. 2021;131(15):148272.
- <span id="page-14-27"></span>29. Kaminiów K, Pająk M, Pająk R, Paprocka J. Pyridoxine-dependent epilepsy and antiquitin defciency resulting in neonatal-onset refractory seizures. Brain Sci. 2021;12(1):65.
- <span id="page-14-28"></span>30. Rankin PM, Harrison S, Chong WK, Boyd S, Aylett SE. Pyridoxinedependent seizures: a family phenotype that leads to severe cognitive defcits, regardless of treatment regime. Dev Med Child Neurol. 2007;49(4):300–5.
- <span id="page-14-29"></span>31. Nasr E, Mamak E, Feigenbaum A, Donner EJ, Mercimek-Mahmutoglu S. Long-term treatment outcome of two patients with pyridoxinedependent epilepsy caused by ALDH7A1 mutations: normal neurocognitive outcome. J Child Neurol. 2015;30(5):648–53.
- <span id="page-14-30"></span>32. Sauer SW, Opp S, Hofmann GF, Koeller DM, Okun JG, Kölker S. Therapeutic modulation of cerebral L-lysine metabolism in a mouse model for glutaric aciduria type I. Brain J Neurol. 2011;134(Pt 1):157–70.
- <span id="page-14-31"></span>33. Gallagher RC, Van Hove JLK, Scharer G, Hyland K, Plecko B, Waters PJ, et al. Folinic acid-responsive seizures are identical to pyridoxinedependent epilepsy. Ann Neurol. 2009;65(5):550–6.
- <span id="page-15-0"></span>34. Chi W, Iyengar ASR, Fu W, Liu W, Berg AE, Wu CF, et al. Drosophila carrying epilepsy-associated variants in the vitamin B6 metabolism gene PNPO display allele- and diet-dependent phenotypes. Proc Natl Acad Sci U S A. 2022;119(9):e2115524119.
- <span id="page-15-1"></span>35. Mills PB, Camuzeaux SSM, Footitt EJ, Mills KA, Gissen P, Fisher L, et al. Epilepsy due to PNPO mutations: genotype, environment and treatment afect presentation and outcome. Brain J Neurol. 2014;137(Pt 5):1350–60.
- <span id="page-15-2"></span>36. di Salvo ML, Mastrangelo M, Nogués I, Tolve M, Paiardini A, Carducci C, et al. Pyridoxine-5'-phosphate oxidase (Pnpo) defciency: Clinical and biochemical alterations associated with the C.347g>A (P.·Arg116gln) mutation. Mol Genet Metab. 2017;122(1–2):135–42.
- <span id="page-15-3"></span>37. Alghamdi M, Bashiri FA, Abdelhakim M, Adly N, Jamjoom DZ, Sumaily KM, et al. Phenotypic and molecular spectrum of pyridoxamine-5'-phosphate oxidase defciency: a scoping review of 87 cases of pyridoxamine-5'-phosphate oxidase defciency. Clin Genet. 2021;99(1):99–110.
- <span id="page-15-4"></span>38. Whyte MP, Zhang F, Wenkert D, McAlister WH, Mack KE, Benigno MC, et al. Hypophosphatasia: validation and expansion of the clinical nosology for children from 25 years experience with 173 pediatric patients. Bone. 2015;75:229–39.
- <span id="page-15-5"></span>39. Mumm S, Jones J, Finnegan P, Whyte MP. Historical vignette: hypophosphatasia: molecular diagnosis of Rathbun's original case. J Bone Miner Res. 2001;16(9):1724–7.
- <span id="page-15-6"></span>40. Whyte MP, Essmyer K, Geimer M, Mumm S. Homozygosity for TNSALP mutation 1348c>T (Arg433Cys) causes infantile hypophosphatasia manifesting transient disease correction and variably lethal outcome in a kindred of black ancestry. J Pediatr. 2006;148(6):753–8.
- <span id="page-15-11"></span>41. Fraser D. Hypophosphatasia. Am J Med. 1957年5月;22(5):730–46.
- <span id="page-15-7"></span>42. Whyte MP. Hypophosphatasia — aetiology, nosology, pathogenesis, diagnosis and treatment. Nat Rev Endocrinol. 2016;12(4):233–46.
- <span id="page-15-8"></span>43. Robison R. The possible signifcance of hexosephosphoric esters in ossifcation. Biochem J. 1923;17(2):286–93.
- <span id="page-15-17"></span>44. Whyte MP. Physiological role of alkaline phosphatase explored in hypophosphatasia. Ann N Y Acad Sci. 2010;1192:190–200.
- <span id="page-15-9"></span>45. Moss DW, Eaton RH, Smith JK, Whitby LG. Association of inorganicpyrophosphatase activity with human alkaline-phosphatase preparations. Biochem J. 1967;102(1):53–7.
- <span id="page-15-10"></span>46. Anderson HC, Hsu HH, Morris DC, Fedde KN, Whyte MP. Matrix vesicles in osteomalacic hypophosphatasia bone contain apatite-like mineral crystals. Am J Pathol. 1997;151(6):1555–61.
- <span id="page-15-12"></span>47. Fedde KN, Blair L, Silverstein J, Coburn SP, Ryan LM, Weinstein RS, et al. Alkaline phosphatase knock-out mice recapitulate the metabolic and skeletal defects of infantile hypophosphatasia. J Bone Miner Res. 1999;14(12):2015–26.
- <span id="page-15-13"></span>48. Gospe SM. Neonatal vitamin-responsive epileptic encephalopathies. Chang Gung Med J. 2010;33(1):1–12.
- <span id="page-15-14"></span>49. Cahill RA, Wenkert D, Perlman SA, Steele A, Coburn SP, McAlister WH, et al. Infantile hypophosphatasia: transplantation therapy trial using bone fragments and cultured osteoblasts. J Clin Endocrinol Metab. 2007;92(8):2923–30.
- 50. Shohat M, Rimoin DL, Gruber HE, Lachman RS. Perinatal lethal hypophosphatasia; clinical, radiologic and morphologic fndings. Pediatr Radiol. 1991;21(6):421–7.
- 51. Silver MM, Vilos GA, Milne KJ. Pulmonary hypoplasia in neonatal hypophosphatasia. Pediatr Pathol. 1988;8(5):483–93.
- <span id="page-15-15"></span>52. Kozlowski K, Sutclife J, Barylak A, Harrington G, Kemperdick H, Nolte K, et al. Hypophosphatasia. Review of 24 cases. Pediatr Radiol. 1976;5(2):103–17.
- <span id="page-15-16"></span>53. Barcia JP, Strife CF, Langman CB. Infantile hypophosphatasia: treatment options to control hypercalcemia, hypercalciuria, and chronic bone demineralization. J Pediatr. 1997;130(5):825–8.
- <span id="page-15-18"></span>54. Belachew D, Kazmerski T, Libman I, Goldstein AC, Stevens ST, DeWard S, et al. Infantile Hypophosphatasia Secondary to a Novel Compound Heterozygous Mutation Presenting with Pyridoxine-Responsive Seizures. Berlin: Springer Berlin Heidelberg; 2013.17–24.
- <span id="page-15-19"></span>55. Whyte MP, Kurtzberg J, McAlister WH, Mumm S, Podgornik MN, Coburn SP, et al. Marrow cell transplantation for infantile hypophosphatasia. J Bone Miner Res Off J Am Soc Bone Miner Res. 2003;18(4):624-36.
- <span id="page-15-20"></span>56. Zeng WQ, Al-Yamani E, Acierno JS, Slaugenhaupt S, Gillis T, MacDonald ME, et al. Biotin-responsive basal ganglia disease maps to 2q36.3 and is due to mutations in SLC19A3. Am J Hum Genet. 2005;77(1):16–26.
- 57. Rajgopal A, Edmondnson A, Goldman ID, Zhao R. SLC19A3 encodes a second thiamine transporter ThTr2. Biochim Biophys Acta. 2001;1537(3):175–8.
- <span id="page-15-21"></span>58. Vlasova TI, Stratton SL, Wells AM, Mock NI, Mock DM. Biotin defciency reduces expression of SLC19A3, a potential biotin transporter, in leukocytes from human blood. J Nutr. 2005;135(1):42–7.
- <span id="page-15-22"></span>59. Pérez-Dueñas B, Serrano M, Rebollo M, Muchart J, Gargallo E, Dupuits C, et al. Reversible lactic acidosis in a newborn with thiamine transporter-2 defciency. Pediatrics. 2013;131(5):e1670-1675.
- <span id="page-15-24"></span>60. Ozand P. Biotin-responsive basal ganglia disease: a novel entity. Brain. 1998;121(7):1267–79.
- <span id="page-15-23"></span>61. Kono S, Miyajima H, Yoshida K, Togawa A, Shirakawa K, Suzuki H. Mutations in a thiamine-transporter gene and Wernicke's-like encephalopathy. N Engl J Med. 2009;360(17):1792–4.
- <span id="page-15-25"></span>62. Eichler FS, Swoboda KJ, Hunt AL, Cestari DM, Rapalino O. Case 38–2017. A 20-year-old woman with seizures and progressive dystonia. N Engl J Med. 2017;377(24):2376–85.
- <span id="page-15-26"></span>63. Yamada K, Miura K, Hara K, Suzuki M, Nakanishi K, Kumagai T, et al. A wide spectrum of clinical and brain MRI fndings in patients with SLC19A3 mutations. BMC Med Genet. 2010;11:171.
- 64. Whitford W, Hawkins I, Glamuzina E, Wilson F, Marshall A, Ashton F, et al. Compound heterozygous SLC19A3 mutations further refne the critical promoter region for biotin-thiamine-responsive basal ganglia disease. Cold Spring Harb Mol Case Stud. 2017;3(6):a001909.
- <span id="page-15-27"></span>65. Bubshait DK, Rashid A, Al-Owain MA, Sulaiman RA. Depression in adult patients with biotin responsive basal ganglia disease. Drug Discov Ther. 2016;10(4):223–5.
- <span id="page-15-28"></span>66. Bin Saeedan M, Dogar MA. Teaching NeuroImages: MRI fndings of biotin-responsive basal ganglia disease before and after treatment. Neurology. 2016;86(7):e71-72.
- <span id="page-15-29"></span>67. Alfadhel M, Almuntashri M, Jadah RH, Bashiri FA, Al Rifai M, Al Shalaan H, et al. Biotin-responsive basal ganglia disease should be renamed biotinthiamine-responsive basal ganglia disease: a retrospective review of the clinical, radiological and molecular fndings of 18 new cases. Orphanet J Rare Dis. 2013;8(1):83.
- <span id="page-15-30"></span>68. Gowda VK, Srinivasan VM, Bhat M, Benakappa N. Biotin thiamin responsive basal ganglia disease in siblings. Indian J Pediatr. 2018;85(2):155–7.
- <span id="page-15-31"></span>69. Chuanjie W, Haiyue Z, Haiqing S, Yuping W, Xunming J. Research Progress on Biotin-Thioammonium Reactive Basal Gangliopathy - Chinese Medical Journal. Natl Med J China. 2018;98(33):2688–90.
- <span id="page-15-32"></span>70. Wolf B. Worldwide survey of neonatal screening for biotinidase defciency. J Inherit Metab Dis. 1991;14(6):923–7.
- <span id="page-15-33"></span>71. Wolf B, Grier RE, Allen RJ, Goodman SI, Kien CL. Biotinidase defciency: the enzymatic defect in late-onset multiple carboxylase defciency. Clin Chim Acta Int J Clin Chem. 1983;131(3):273–81.
- <span id="page-15-34"></span>72. Micó SI, Jiménez RD, Salcedo EM, Martínez HA, Mira AP, Fernández CC. Epilepsy in biotinidase defciency after biotin treatment. JIMD Rep. 2012;4:75–8.
- <span id="page-15-35"></span>73. Bunch M, Singh A. Peculiar neuroimaging and electrophysiological fndings in a patient with biotinidase defciency. Seizure. 2011;20(1):83–6.
- <span id="page-15-36"></span>74. Desai S, Ganesan K, Hegde A. Biotinidase defciency: a reversible metabolic encephalopathy. Neuroimaging and MR spectroscopic fndings in a series of four patients. Pediatr Radiol. 2008;38(8):848–56.
- <span id="page-15-37"></span>75. Singhi P, Ray M. Ohtahara syndrome with biotinidase defciency. J Child Neurol. 2011;26(4):507–9.
- <span id="page-15-38"></span>76. Mock DM. Skin manifestations of biotin defciency. Semin Dermatol. 1991;10(4):296–302.
- <span id="page-15-39"></span>77. Navarro PC, Guerra A, Alvarez JG, Ortiz FJ. Cutaneous and neurologic manifestations of biotinidase defciency. Int J Dermatol. 2000;39(5):363–5.
- <span id="page-15-40"></span>78. Schürmann M, Engelbrecht V, Lohmeier K, Lenard HG, Wendel U, Gärtner J. Cerebral metabolic changes in biotinidase defciency. J Inherit Metab Dis. 1997;20(6):755–60.
- <span id="page-15-41"></span>79. Micó SI, Jiménez RD, Salcedo EM, Martínez HA, Mira AP, Fernández CC. Epilepsy in Biotinidase Defciency After Biotin Treatment. Berlin: Springer Berlin Heidelberg; 2011. 75–8.
- <span id="page-15-42"></span>80. Yang Y, Li C, Qi Z, Xiao J, Zhang Y, Yamaguchi S, et al. Spinal cord demyelination associated with biotinidase defciency in 3 Chinese patients. J Child Neurol. 2007;22(2):156–60.
- <span id="page-15-43"></span>81. Salbert BA, Pellock JM, Wolf B. Characterization of seizures associated with biotinidase defciency. Neurology. 1993;43(7):1351–5.
- <span id="page-16-0"></span>82. Schulpis KH, Karikas GA, Tjamouranis J, Regoutas S, Tsakiris S. Low serum biotinidase activity in children with valproic acid monotherapy. Epilepsia. 2001;42(10):1359–62.
- <span id="page-16-1"></span>83. Bhat AS, Srinivasan K, Kurpad SS, Galgali RB. Psychiatric presentations of vitamin B 12 defciency. J Indian Med Assoc. 2007;105(7):395–6.
- <span id="page-16-2"></span>84. Vilibić M, Jukić V, Vidović A, Brecić P. Cobalamin defciency manifested with seizures, mood oscillations, psychotic features and reversible dementia in the absence of typical neurologic and hematologic signs and symptoms: a case report. Coll Antropol. 2013;37(1):317–9.
- <span id="page-16-3"></span>85. Gramer G, Fang-Hoffmann J, Feyh P, Klinke G, Monostori P, Mütze U, et al. Newborn Screening for Vitamin B12 Defciency in Germany-Strategies, Results, and Public Health Implications. J Pediatr. 2020;216:165-172.e4.
- <span id="page-16-4"></span>86. Irevall T, Axelsson I, Naumburg E. B12 defciency is common in infants and is accompanied by serious neurological symptoms. Acta Paediatr Oslo Nor 1992. 2017;106(1):101–4.
- <span id="page-16-5"></span>87. Dror DK, Allen LH. Effect of vitamin B12 deficiency on neurodevelopment in infants: current knowledge and possible mechanisms. Nutr Rev. 2008;66(5):250–5.
- 88. Akaike A, Tamura Y, Sato Y, Yokota T. Protective effects of a vitamin B12 analog, methylcobalamin, against glutamate cytotoxicity in cultured cortical neurons. Eur J Pharmacol. 1993;241(1):1–6.
- <span id="page-16-6"></span>89. Kumar S. Recurrent seizures: an unusual manifestation of vitamin B12 defciency. Neurol India. 2004;52(1):122–3.
- <span id="page-16-7"></span>90. Silva B, Velosa A, Barahona-Corrêa JB. Reversible dementia, psychotic symptoms and epilepsy in a patient with vitamin  $B_{12}$  deficiency. BMJ Case Rep. 2019;12(5):e229044.
- 91. Naha K, Dasari S, Vivek G, Prabhu M. Vitamin B12 defciency: an unusual cause for recurrent generalised seizures with pancytopaenia. Case Rep. 2012; 2012(aug31 1):bcr2012006632–bcr2012006632.
- 92. Dogan M, Ariyuca S, Peker E, Akbayram S, Dogan ŞZ, Ozdemir O, et al. Psychotic disorder, hypertension and seizures associated with vitamin B12 defciency: a case report. Hum Exp Toxicol. 2012;31(4):410–3.
- <span id="page-16-8"></span>93. Glaser K, Girschick HJ, Schropp C, Speer CP. Psychomotor development following early treatment of severe infantile vitamin B12 defciency and West syndrome – Is everything fne? A case report and review of literature. Brain Dev. 2015;37(3):347–51.
- <span id="page-16-9"></span>94. Malbora B, Yuksel D, Aksoy A, Ozkan M. Two infants with infantile spasms associated with vitamin B12 defciency. Pediatr Neurol. 2014;51(1):144–6.
- <span id="page-16-12"></span>95. Roschitz B, Plecko B, Huemer M, Biebl A, Foerster H, Sperl W. Nutritional infantile vitamin B12 defciency: pathobiochemical considerations in seven patients. Arch Dis Child Fetal Neonatal Ed. 2005;90(3):F281-282.
- <span id="page-16-10"></span>96. Pavone P, Sullo F, Falsaperla R, Greco F, Crespo A, Calvo A, et al. Vitamin B12 defciency and west syndrome: an uncommon but preventable cause of neurological disorder. Report on three cases, one of them with late onset during vitamin B12 treatment. Neuropediatrics. 2021;52(04):333–6.
- <span id="page-16-11"></span>97. Meena MK, Sharma S, Bhasin H, Jain P, Kapoor S, Jain A, et al. Vitamin  $B_{12}$  deficiency in children with infantile spasms: a case-control study. J Child Neurol. 2018;33(12):767–71.
- <span id="page-16-13"></span>98. Karabiber H, Sonmezgoz E, Ozerol E, Yakinci C, Otlu B, Yologlu S. Efects of valproate and carbamazepine on serum levels of homocysteine, vitamin B12, and folic acid. Brain Dev. 2003;25(2):113–5.
- <span id="page-16-14"></span>99. Schwahn BC, Chen Z, Laryea MD, Wendel U, Lussier-Cacan S, Genest J, et al. Homocysteine-betaine interactions in a murine model of 5,10-methylenetetrahydrofolate reductase defciency. FASEB J Of Publ Fed Am Soc Exp Biol. 2003;17(3):512–4.
- <span id="page-16-15"></span>100. Steinfeld R, Grapp M, Kraetzner R, Dreha-Kulaczewski S, Helms G, Dechent P, et al. Folate receptor alpha defect causes cerebral folate transport defciency: a treatable neurodegenerative disorder associated with disturbed myelin metabolism. Am J Hum Genet. 2009;85(3):354–63.
- <span id="page-16-16"></span>101. Ramaekers VT, Blau N. Cerebral folate defciency. Dev Med Child Neurol. 2004;46(12):843–51.
- <span id="page-16-17"></span>102. Cario H, Bode H, Debatin KM, Opladen T, Schwarz K. Congenital null mutations of the FOLR1 gene: a progressive neurologic disease and its treatment. Neurology. 2009;73(24):2127–9.
- 103. Al-Baradie RS, Chaudhary MW. Diagnosis and management of cerebral folate defciency. A form of folinic acid-responsive seizures. Neurosci Riyadh Saudi Arab. 2014;19(4):312–6.
- 104. Toelle SP, Wille D, Schmitt B, Scheer I, Thöny B, Plecko B. Sensory stimulus-sensitive drop attacks and basal ganglia calcifcation: new

fndings in a patient with FOLR1 defciency. Epileptic Disord Int Epilepsy J Videotape. 2014;16(1):88–92.

- 105. Delmelle F, Thöny B, Clapuyt P, Blau N, Nassogne MC. Neurological improvement following intravenous high-dose folinic acid for cerebral folate transporter defciency caused by FOLR-1 mutation. Eur J Paediatr Neurol EJPN Off J Eur Paediatr Neurol Soc. 2016;20(5):709–13.
- 106. Karin I, Borggraefe I, Catarino CB, Kuhm C, Hoertnagel K, Biskup S, et al. Folinic acid therapy in cerebral folate defciency: marked improvement in an adult patient. J Neurol. 2017;264(3):578–82.
- 107. Kobayashi Y, Tohyama J, Akiyama T, Magara S, Kawashima H, Akasaka N, et al. Severe leukoencephalopathy with cortical involvement and peripheral neuropathy due to FOLR1 defciency. Brain Dev. 2017;39(3):266–70.
- <span id="page-16-18"></span>108. Pérez-Dueñas B, Toma C, Ormazábal A, Muchart J, Sanmartí F, Bombau G, et al. Progressive ataxia and myoclonic epilepsy in a patient with a homozygous mutation in the FOLR1 gene. J Inherit Metab Dis. 2010;33(6):795–802.
- <span id="page-16-19"></span>109. Bonkowsky JL, Ramaekers VT, Quadros EV, Lloyd M. Progressive Encephalopathy in a Child with Cerebral Folate Defciency Syndrome. J Child Neurol. 2008;23(12):1460–3.
- <span id="page-16-20"></span>110. Maf S, Laroche-Raynaud C, Chazelas P, Lia AS, Derouault P, Sturtz F, et al. Pharmacoresistant epilepsy in childhood: think of the cerebral folate defciency, a treatable disease. Brain Sci. 2020;10(11):762.
- <span id="page-16-21"></span>111. Papadopoulou MT, Dalpa E, Portokalas M, Katsanika I, Tirothoulaki K, Spilioti M, et al. Cerebral folate defciency in two siblings caused by biallelic variants including a novel mutation of *FOLR1* gene: Intrafamilial heterogeneity following early treatment and the role of ketogenic diet. JIMD Rep. 2021;60(1):3–9.
- <span id="page-16-22"></span>112. Qiu A, Jansen M, Sakaris A, Min SH, Chattopadhyay S, Tsai E, et al. Identifcation of an intestinal folate transporter and the molecular basis for hereditary folate malabsorption. Cell. 2006;127(5):917–28.
- <span id="page-16-23"></span>113. Nakai Y, Inoue K, Abe N, Hatakeyama M, Ohta KY, Otagiri M, et al. Functional characterization of human proton-coupled folate transporter/heme carrier protein 1 heterologously expressed in mammalian cells as a folate transporter. J Pharmacol Exp Ther. 2007;322(2):469–76.
- <span id="page-16-24"></span>114. Geller J, Kronn D, Jayabose S, Sandoval C. Hereditary folate malabsorption: family report and review of the literature. Medicine (Baltimore). 2002;81(1):51–68.
- <span id="page-16-25"></span>115. Allen RH, Stabler SP, Lindenbaum J. Serum betaine, N, N-dimethylglycine and N-methylglycine levels in patients with cobalamin and folate defciency and related inborn errors of metabolism. Metabolism. 1993;42(11):1448–60.
- <span id="page-16-26"></span>116. Cario H, Smith DEC, Blom H, Blau N, Bode H, Holzmann K, et al. Dihydrofolate reductase defciency due to a homozygous dhfr mutation causes megaloblastic anemia and cerebral folate defciency leading to severe neurologic disease. Am J Hum Genet. 2011;88(2):226–31.
- <span id="page-16-27"></span>117. Banka S, Blom HJ, Walter J, Aziz M, Urquhart J, Clouthier CM, et al. Identifcation and characterization of an inborn error of metabolism caused by dihydrofolate reductase defciency. Am J Hum Genet. 2011;88(2):216–25.
- <span id="page-16-28"></span>118. Ponzone A, Spada M, Ferraris S, Dianzani I, de Sanctis L. Dihydropteridine reductase defciency in man: from biology to treatment. Med Res Rev. 2004;24(2):127–50.
- <span id="page-16-29"></span>119. Pollock RJ, Kaufman S. Dihydropteridine reductase may function in tetrahydrofolate metabolism. J Neurochem. 1978;31(1):115–23.
- <span id="page-16-30"></span>120. de Koning TJ, Duran M, Dorland L, Gooskens R, Van Schaftingen E, Jaeken J, et al. Benefcial efects of L-serine and glycine in the management of seizures in 3-phosphoglycerate dehydrogenase defciency. Ann Neurol. 1998;44(2):261–5.
- <span id="page-16-31"></span>121. Poli A, Vial Y, Haye D, Passemard S, Schiff M, Nasser H, et al. Phosphoglycerate dehydrogenase (PHGDH) defciency without epilepsy mimicking primary microcephaly. Am J Med Genet A. 2017;173(7):1936–42.
- <span id="page-16-32"></span>122. Jaeken J, Detheux M, Van Maldergem L, Foulon M, Carchon H, Van Schaftingen E. 3-Phosphoglycerate dehydrogenase defciency: an inborn error of serine biosynthesis. Arch Dis Child. 1996;74(6):542–5.